

Environmental Prevalence of Pathogens in Different Drinking Water Resources in Makkah City (Kingdom of Saudi Arabia)

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ABSTRACT

Water is the most important substance in our daily life. Without it, life would not have been possible. Potable water is essential to humans and other life forms, as water is important to the mechanics of biological metabolisms in the body. Drinking water should be pure and free of contaminants to ensure proper health and wellness. Drinking water from different water resources such as wells and tankers should be free from contamination with waterborne pathogens including bacteria, fungi, viruses and parasites. Treatment of water using many ways is generally done in order to purify it. However, some water treatment techniques may not properly handled. In addition, water transferring techniques may contaminate the drinking water. Therefore, this study was aimed to investigate drinking water in wells and tankers to observe any microbial pathogen presence as a source of health hazard. One hundred and eight water samples from different sources were examined for microbial pathogens using filtration method on solid and liquid selective media. Four sources include sea desalinated water (SDW) from governmental water desalination factories, drinkable wells water (DWW), non-drinkable wells water (NDWW) and commercial desalinated water (CDW) from small commercial water desalination factories. Seven DWW samples (58.3%) and five NDWW samples (41.7%) were contaminated with *E. coli*. Eleven DWW samples (91.7%) and all NDWW samples (100%) were contaminated with *P. aeruginosa*. One DWW sample (8.3%) and two NDWW samples (16.7%) were contaminated with *E. faecalis*. Four DWW samples (33.3%) and one NDWW sample (8.3%) were found contaminated with *Aspergillus* spp. Four SDW samples (100%) and four CDW samples (50%) were contaminated with *Penicillium* spp. Conclusion: CDW was found to be the more suitable than other sources for drinking if a biological hazard is the main target. However, contamination at transferring process should be addressed. Yet, water tanker which is a common transferring technique in many areas in Saudi Arabia and should be tested for safety level from point of contamination hazard during the transferring process.

INTRODUCTION

Water is unsafe for human consumption when it contains pathogenic or disease-causing microorganisms. Pathogenic microorganisms (and their associated disease(s)) may include bacteria, such as *Salmonella typhi* (typhoid fever), *Vibrio cholerae* (cholera), *Shigella* (dysentery, shigellosis), and viruses, such as poliovirus or Hepatitis A virus and protozoa such as *Giardia lamblia* (giardiasis) or *Cryptosporidium parvum* (cryptosporidiosis). A major challenge for water suppliers is how to control and limit the risks from pathogens and disinfection by-products. It is

important to provide protection from pathogens while simultaneously minimizing health risks to the population from disinfection by-products (EPA, 2011).

In addition to bacterial-related health risks, faecal contamination carries the increased risk of viral contamination of the water source. Although viruses cannot multiply in water, some may remain static. This health risk is elevated in treated water (i.e. chlorinated tanks) where the faecal indicators may be absent. Many viruses have been identified as key etiological agents in outbreaks of drinking water derived gastrointestinal illness in the United States and Netherlands (Leclerc *et al.*, 2002).

Some micro-fungi are known to be opportunistic human pathogens. Airborne spores are an important potential source of microfungi found in water storage reservoirs. It has also demonstrated conclusively that filamentous microfungi grow and sporulate on the inner surfaces of water pipe and in soft sediments within the water distribution system (Sammon *et al.*, 2011).

Worldwide, over one billion people lack access to an adequate water supply; more than twice as many lack basic sanitation (WHO/UNICEF, 2006). Unsafe water, inadequate sanitation, and insufficient hygiene account for an estimated 9.1 percent of the global burden of disease and 6.3 percent of all deaths, according to the World Health Organization (Prüss-Ustün *et al.*, 2008). This burden is disproportionately borne by children in developing countries, with water-related factors causing more than 20 percent of deaths of people under age 14. Nearly half of all people in developing countries have infections or diseases associated with inadequate water supply and sanitation (Bartram *et al.*, 2005).

The presence of *E.coli* in water is a strong indication of recent sewage or faecal contamination. Sewage may contain many types of disease-causing organisms. *E. coli* comes from human and animal waste. During rainfalls, snow melts, or other types of precipitation, *E.coli* may be washed into creeks, rivers, streams, lakes, or groundwater. When these waters are used as sources of drinking water and the water is not treated or inadequately treated, *E.coli* may end up in the drinking water (Health Canada, 2008).

Faecal coliforms and *E.coli* are bacteria whose presence indicates that the water may be contaminated with human or animal wastes. Microbes in these waters can cause short-term effects, such as diarrhea, cramps, nausea, headaches, or other symptoms. They may pose a special health risk for infants, young children, some of the elderly, and people with severely compromised immune systems (CDC, 2009).

Some bacteria are ubiquitous in soil, water and on surfaces in contact with soil or water such as *Pseudomonas aeruginosa* which is

an opportunistic pathogen. *P.aeruginosa* is an opportunistic pathogen. It produces tissue-damaging toxins and causes urinary tract infections, respiratory system infections, central nervous system, endocarditis (*P.aeruginosa* infects heart valves establishes itself on the endocardium), dermatitis, soft tissue infections, bacteraemia, bone and joint infections, gastrointestinal infections and a variety of systemic infections, particularly in patients with severe burns and in cancer and AIDS patients who are immunosuppressed (EHA, 2012).

Spread occurs from patient to patient on the hands of hospital personnel, by direct patient contact with contaminated reservoirs, and by the ingestion of contaminated foods and water (EHA, 2012).

The presence of faecal coliform in aquatic environments may indicate that the water has been contaminated with the faecal material of humans or animals. Faecal coliform bacteria can enter water bodies through direct discharge of waste from mammals and birds, from storm and agricultural runoff, and from human waste (Doyle and Erickson, 2006).

Pet wastes (cats, dogs) can contribute to faecal contamination of surface waters. Runoff from roads, parking lots, and yards can carry animal wastes to streams through storm sewers. Birds can be a significant source of faecal coliform bacteria. Birds (seagulls, geese, swans) can all elevate bacterial counts, especially in freshwater systems (wetland, rivers, lakes and ponds). Some waterborne pathogenic diseases that may coincide with faecal coliform contamination include ear infections, viral and bacterial gastroenteritis, dysentery, typhoid fever and hepatitis A. The presence of faecal coliform tends to affect humans more than it does aquatic creatures, though not exclusively (Walkerton, 2011).

Fungi are ubiquitous organisms that are widely distributed in nature. Several fungal genera have been shown to be allergenic, such as *Aspergillus*, *Alternaria* and *Cladosporium* (Black *et al.*, 2000; Bowyer *et al.*, 2006; Hedayati *et al.*, 2007; Simon-Nobbe *et al.*, 2006). Several studies have suggested an important role for waterborne

fungi to endanger human health (Anaissie *et al.*, 2001; 2003; Warris *et al.*, 2001). Some of these studies have linked a genetic relationship between waterborne fungi and fungi isolated from clinical samples (Anaissie *et al.*, 2001; 2003).

There are several species of *Aspergillus* which cause infection to the human especially *Aspergillus fumigatus*. Aspergillosis is opportunistic respiratory infection which causes about 40% of fatal nosocomial infections. *Aspergillus* spp infections are transmitted by water (Graybill, 2001). Drinking water quality is usually determined by its pathogenic bacterial content. However, the potential of water-borne spores as a source of nosocomial fungal infection is increasingly being recognized. Sammon *et al.*, (2010) demonstrated that numerous microfungus genera, including those that contain species which are opportunistic human pathogens, populate a typical treated municipal water supply in sub-tropical Australia.

Penicillium spp. contain more than 225 species confirming to certain morphological criteria (Pitt *et al.*, 2000). They can be isolated from the well water (Siqueira *et al.*, 2011). *Penicillium marneffi* is dimorphic, forming yeast-like cells in infected tissues, often they can be found intracellular (Jolanta, 2005; Rajendran *et al.*, 2006). In immunocompromised people, *P. marneffi* considered as a common opportunistic pathogen which can cause systemic penicillosis in acquired immunodeficiency syndrome (AIDS) patients. Rodents are usual reservoirs for *P. marneffi* and may be involved in its transmission to humans (Cheesbrough, 2007).

Pathogenic bacteria can occur in surface water in large numbers, either being excreted in faeces or occurring naturally in the environment. Bacteria typically range in size between 0.5 and 2 micrometres. Disease-causing bacteria that can be transmitted by water include *Vibrio cholerae*, *Salmonella* sp, *Campylobacter* sp, *Shigella* sp, and *Staphylococcus aureus*. (Health Canada, 2006a).

Aims of the present study was to investigate drinking water in wells and tankers, and discover any microbial pathogens in these water as a source of biological environmental health hazard.

MATERIALS AND METHODS

Sources of water samples

One hundred and eight water samples (36 samples in triplicate) were collected from different water sources within Makkah city and analysed for bacterial and fungal contamination. Each sample was collected in sterile container sealed with screw cap after disinfection of dispensing point with flame. Then, samples were kept on ice till analysis take place in the laboratory within three hours. There were four sources of water included in this study: governmental sea desalinated water (12 samples), drinkable wells water (36 samples), non-drinkable wells water (36 samples) and small commercial desalination water factories (24 samples).

Sample analysis

Each sample was diluted with sterile distilled water at ratio of 1:10 as final volume 300 ml. Then, each 100 ml from the diluted sample was filtered using filtration equipment system (LabTech, Korea) with fresh cellulose nitrate filter (Sartorius, Germany, with pore size 0.45 µm) for each partition of the diluted sample.

The three partitions were poured through filter trap, then two cellulose nitrate filter was taken out carefully by sterile forceps and placed on the MacConkey plate (Biolab, Hungary) and bile esculin plate (Himedia, India) for bacterial growth and the third filter was placed on SD media plate (Himedia, India) for fungal growth. MacConkey and bile esculin plates were incubated for 24 hrs °C, while SD plate at 25 °C for 72 hrs (Harley *et al.*, 2002).

Isolation of microorganisms in water samples

Bacterial identification

Colony counter was used for counting of bacterial colonies on cellulose nitrate filters. The used formula for calculating number of bacteria per 100 ml water sample is:

$$\text{No. of bacteria/100 ml} = \text{colony dilution} \times \text{dilution factor.}$$

Cellulose nitrate filter was placed and cultured in MacConkey or bile esculine media.

Table 1: The percentages of bacterial contamination in investigated water samples

Bacteria	Drinkable well water (n=36)		Non-drinkable well water (n=36)		Desalinated water (n=12)		Private desalinated water (n=24)	
	No.	%	No.	%	No.	%	No.	%
<i>E. coli</i>	21	58.3	15	41.7	0	0	0	0
<i>P. aeruginosa</i>	33	91.7	36	100	0	0	0	0
<i>E. faecalis</i>	3	8.3	6	16.7	0	0	0	0

n = Means total number of investigated samples from each water source.

No. = Means number of positively contaminated water samples

Table 2: The percentages of fungal contamination in investigated water samples

Bacteria	Drinkable well water (n=36)		Non-drinkable well water (n=36)		Desalinated water (n=12)		Private desalinated water (n=24)	
	No.	%	No.	%	No.	%	No.	%
<i>Aspergillus</i> spp.	12	33.3	3	8.3	0	0	0	0
<i>Penicillium</i> spp	0	0	0	0	12	100	12	50

n = Means total number of investigated samples from each water source.

No. = Means number of positively contaminated water samples

Table 3: Isolated *E. coli* from water samples

Type of water/location	Source of water	<i>E. coli</i>	
		Mean Colony count n =3	No. of bacteria/ 100 ml
Drinkable wells water	Well 46	WT 1	30
		WT 2	50
		WT 3	40
	Well 48	WS	20
		WT 1	20
		WT 2	40
Non-drinkable wells water	Well 27	WT 3	10
		WT 1	10
		WT 3	20
	Well 47	WT 1	10
		WT 2	30
	Well 52	WT 2	20

WS: water source, WT: Water Tanker

Colonies in MacConkey media were pink or yellow color, small size, while colonies in bile esculine media were black in color. On MacConkey media, pink colonies indicate lactose fermented bacteria like *E. coli* while yellow colonies indicate non lactose fermented bacteria like *P. aeruginosa*. The non-lactose fermented bacteria were cultured on nutrient agar media to confirm *P. aeruginosa* that give greenish color colonies. The black colonies in bile esculine media indicate *E. faecalis*.

Gram stain

One drop of saline was mixed with a single colony on slide and fixed with gentle heat. Crystal Violet Oxalate (Atlas, UK) was poured on slide for 2-3 minutes. Gram's Iodine (mordant) was poured on slide for 2-3 minutes. Alcohol decolorized was poured on slide for 1 minute. Safranin counterstain was poured on slide for 2-3 minutes. In each step, the slide was washed with distilled water. The slide

Table 4: Isolated *P. aeruginosa* from water samples

Type of water/location	Source of water	<i>P. aeruginosa</i>			
		Mean Colony count n =3	No. of bacteria/ 100 ml		
Drinkable wells water	Well 46	WT 1	37	370	
		WT 2	30	300	
		WT 3	12	120	
	Well 48	WS	103	1030	
		WT 1	74	740	
		WT 2	47	470	
		WT 3	22	220	
		Well 233	WS	6	60
			WT 1	26	260
	WT 2		11	110	
	Non-drinkable wells water	Well 27	WT 3	17	170
			WS	2	20
WT 1			22	220	
Well 47		WT 2	15	150	
		WT 3	8	80	
		WS	3	30	
		WT 1	117	1170	
		WT 2	23	230	
		WT 3	4	40	
Well 52		WS	5	50	
		WT 1	106	1060	
		WT 2	5	50	
		WT 3	7	70	

WS: water source, WT: Water Tanker

Table 5: Isolated *E. faecalis* from water samples

Type of water/location	Source of water	<i>E. faecalis</i>		
		Mean Colony count N=3	No. of bacteria/100 ml	
Drinkable wells water	Well 46	WT 2	10	100
Non-drinkable wells water	Well 47	WT 1	20	200
		WT 2	50	500

WS: water source, WT: Water Tanker

was examined microscopically according to Cheesbrough (2007). Gram positive bacteria were blue or violet while gram negative bacteria were pink or red (Momenah, 2004).

Brilliant Green Lactose Bile (BGLB) tube test

Every pink colony from MacConkey media was cultured on BGLB (Biolab, Hungary) tube containing inverted Durham's tube. The tube was incubated for 48 hours at 44 C to detect *E. coli*. The formation of gas in Durham's tube was recorded if the sample was positive (Collee *et al.*, 1989).

Indole test

Used to identify *E. coli* (indole positive). Each pink colonies from MacConkey media was cultured into tube of tryptone water (HIMEDIA, India). The tube incubated at 44 C. A few drops of Kovak's reagent (BioMerieux, France) added the tryptone water culture after the incubation. After gently mixing of tube, positive indole test was indicated by formation of red color in the surface layer within 10 minutes Cheesbrough (2007).

Oxidase test

Used to identify *P. aeruginosa* (oxidase positive). 2-3 drops of freshly prepared oxidase reagent (BDH Laboratories, UK) were added to a piece of filter paper, then the plastic loop was used to take out a colony of the organism that appeared in the culture and placed on the filter paper. In positive cases, blue-purple color appears in few seconds Cheesbrough (2007).

Fungi identification

Macroscopic examination

On SD media, texture, surface color and pigment of the reverse (underside) appeared in positive fungal growth. Fungi can cover the whole surface of SD media.

Microscopic examination

A small portion of the fungal growth was mixed with drops of Lactophenol Cotton Blue (LPCB)(Bios Europe, UK) on slide. The mixture was tested by using a pair of bent dissecting needles and the slide placed with cover. The cover slip pressed softly by using eraser end of the pencil. The slide tested by low and high power for presence of macroconidia, microconidia, spores and hyphae.

RESULTS

Seven drinkable well water samples (58.3%) and five non-drinkable well water samples (41.7%) were contaminated with *E. coli* (tables 1,3). Eleven drinkable well water samples (91.7%) and all non-drinkable well water samples (100%) were contaminated with *P. aeruginosa*(tables 1,4). One drinkable well water samples (8.3%) and 2 non-drinkable well water samples (16.7%) were contaminated with *E. faecalis* (tables 1,5). Four

Table 6: Isolated *Aspergillus* spp from water samples

Type of water/location	Source of water	
Drinkable wells water	Well 46	WT 2
	Well 48	WT 3
	Well 233	WS WT 3
Non-drinkable wells water	Well 52	WT 3

Table 7: Isolated *Penicillium* spp. from water samples

Type of water/location	Source of water		
Drinkable wells water	Al-Abdiah	WS WT 1 WT 2 WT 3	
		Al-Shalal Al-Aziziah Al-Kadir Qatrat Al-Nadah	Before filtration

WS: water source

WT: Water Tanker

drinkable water samples (33.3%) and one non-drinkable water sample (8.3%) were found contaminated with *aspergillus* spp (tables 2,6). Four desalinated water samples (100%) and four private desalinated water samples (50%) were contaminated with *Penicillium* spp (tables 2,7).

DISCUSSION

Waterborne pathogens can cause a problem to drinking water supplies, recreational waters, and source waters for agriculture, and aquaculture. Sources of pathogens include municipal wastewater effluents, urban runoff, agricultural wastes and wildlife. A drinking water killed 54 people and induced illness 400,000 in Milwaukee at 1993. Over 200 outbreaks of infectious diseases in Canada associated with drinking water occurred between 1974-1996. Pathogen contamination of irrigation water or shellfish beds can produce risks to human food supplies. In addition, declines in amphibian populations may be related to fungal or viral pathogens (The National Water Research Institute, Canada, 2001).

The microbiological guidelines and standards for drinking water for *E. coli*, *P. aeruginosa* and *E. faecalis* are zero colony count/100 ml of water sample (The Natural Mineral Water, Spring Water and Bottled Drinking Water Regulations, 1999).

Periodicity and intensity of rainfall have been shown to impact level of microbial contaminant entering tanks, and the more time between events, the more contaminants accumulate and are washed into the tank (Abbott *et al.*, 2006).

Outbreaks of disease attributable to drinking water in USA still occur and can lead to serious acute, chronic, or sometimes fatal health consequences, particularly in sensitive and immunocompromised populations. From 1971 to 2002, there were 764 waterborne outbreaks associated with drinking water, resulting in 575,457 cases of illness and 79 deaths (Blackburn *et al.*, 2004). Contamination of water is affected by the number of pathogens in the source water, the age of the distribution system, the quality of the delivered water, and climatic conditions. Others have recently estimated waterborne illness rates of 12M cases/year and 16 million cases/yr. (Craun *et al.*, 2006). Reynolds *et al.*, (2008) found that 10.7 million infections and 5.4 million illnesses/year occur in populations served by community groundwater systems; 2.2 million infections and 1.1 million illnesses/year occur in noncommunity groundwater

systems; and 26.0 million infections and 13.0 million illnesses/year occur in municipal surface water systems. The total estimated number of waterborne illnesses/yr. in the U.S. is therefore estimated to be 19.5 million/year. The safety of drinking water is evaluated by the results obtained from faecal indicators during the stipulated controls fixed by the legislation. However, drinking-water related illness outbreaks are still occurring worldwide (Figueras and Borrego, 2010).

Drinking-water quality in both urban and rural areas of Pakistan is not being managed properly. Most of the drinking-water supplies are faecally contaminated. At places groundwater quality is deteriorating due to the naturally occurring subsoil contaminants or to anthropogenic activities. The poor bacteriological quality of drinking-water has frequently resulted in high incidence of waterborne diseases while subsoil contaminants have caused other ailments to consumers (Aziz, 2005).

In the developing world, an estimated 10 million young children died there in 2006. Of these deaths, WHO estimates that 16.5 percent, or at least 1.65 million, were due to diarrheal diseases, many of which were caused by contaminated water (WHO, 2008). Deaths caused by nondiarrheal infections like typhoid fever are also related to contaminated water (Crump *et al.*, 2040).

E.coli

The obtained results revealed that *Escherichia Coli* (*E. coli*) was found in water sources and tankers. Some water tankers were contaminated with *E. coli* either due to lack of proper treatment and cleaning of tankers or contamination of water well sources. However, the presence of *E. coli* indicates recent sewage or animal waste contamination (EPA, 2001). *E. coli* contamination in the present study was found to be higher than that recorded by Golas *et al.*, (2002) which represent 10.7% and Malakauskas *et al.*, (2007) found 16.7% of *E. coli* contamination in Lithuania different wells. Admassu *et al.*, (2004) found 28.6% of *E. coli* contamination in wells water while was 58.33% in 12 water samples (Oyetayo *et al.*, 2007) which is higher than that reported in our study.

The survival of enteric bacteria, like *Salmonella spp.* and *E. coli*, within the tank environment is influenced by temperature and the presence of nutrients (Leclerc *et al.*, 2002). While leaves are a common source of organic matter, dust, especially during times of drought when particles of ground-based organic and inorganic matter can be carried for hundreds of kilometres during dust storms (Goudie, 2009), is another source. Evans *et al.*, (2007) demonstrated that airborne pathogens from surrounding soils, including *E. coli*, are significant contributors to the microbial contamination of tanks. Therefore, in addition to the commonly attributed sources of microbial contaminants, birds, possums or rodents defecating or dying in tanks (Australian Government, 2004), rural participants who graze sheep and cattle, may be exposed to microbial contaminants from higher order-mammals. Exposure to microbial contaminants from higher-order mammals brings with it an increased health risk due to the greater zoonotic potential of such microorganisms. The Australian Government in its guidance on use of rainwater tanks (Australian Government, 2004) identifies livestock waste as a health hazard only for underground tanks and aerosol waste appears unconsidered. This partitioning of risk, between in-ground and above-ground tanks needs to be reconsidered in light of the work (Evans *et al.*, 2007).

Increasing outbreaks (Callaway *et al.*, 2009; Goode *et al.*, 2009) of gastrointestinal illness from livestock-derived *E. coli* indicates it is perhaps time to rethink the significance of the presence of *E. coli* in tanks, particularly in rural areas with livestock. In 2006, the most notorious strain of *E. coli*, STEC O157, was responsible for a waterborne-related outbreak that affected more than 100 people across America and caused at least one death when bacteria transferred from a contaminated water source to a spinach crop that was then packaged and widely distributed (Bettelheim 2007).

The survival rate of *E. coli* in water is varies from 13-245 days (LeJeune *et al.*, 2001). Because it can travel long distances underground, it is can be used as indicator for faecal contamination of ground water (Foppen and Schijven, 2006).In

Ireland, bacterial contamination of water is a national concern, with the EPA reporting that over 25% of groundwater samples were contaminated with *E. coli* in 2004 to 2006. *E. coli* is the most important indicator used in Ireland and its presence indicates water is unfit for human consumption. It has long been thought that *E. coli* can only survive for short periods of time in the environment, hence its almost universal use as an indicator of recent faecal contamination of waterways (APA Teagasc, 2010).

E. coli O157:H7 was isolated from many water wells in cattle farms. It may be found in water sources, such as private wells, that have been contaminated with feces from infected humans or animals. Waste can enter the water through: sewage overflows, sewage systems that are not working properly, polluted storm water runoff, and agricultural runoff. Wells may be more vulnerable to such contamination after flooding, particularly if the wells are shallow, have been dug or bored, or have been submerged by floodwater for long periods of time (CDC, 2009).

The Ministry of Health's Annual Report on Drinking-Water in New Zealand showed unacceptable levels of *E. coli* were found in the water of 72,000 or 2 percent of people accessing a registered water supply (Danya, 2011). Parts of the Danish capital Copenhagen were without clean drinking water after high levels of the *E. coli* bacteria were detected in the municipal tap water system (Berlingske daily's Web site, 2011).

Australia is particularly vulnerable to threats to both the quality and quantity of drinking water availability because most rainfall evaporates quickly, resulting in twenty percent of the population relying on ground water for drinking supplies which "is extremely difficult to clean up if it becomes polluted" (NHMRC 2004). More than half of the tank water sampled failed to meet the Australian Drinking Water Guidelines for safe drinking water. Levels of *E. coli* were up to 230x more than the acceptable levels proposed by the Australian Drinking Water Guidelines. Qualitative research found most consumers were unaware of the risks associated with drinking raw rainwater. Further, few took steps to minimize their risk through accepted

water management practices (Andrea and Angela, 2010).

Contamination of water samples with *Pseudomonas aeruginosa*

Our results showed that *Pseudomonas aeruginosa* (PA) was the most common microbial contamination in water sources and tankers. All water tankers and majority of water sources (wells) were contaminated due to lack of proper treatment and cleaning. High numbers of PA which represent 1030 bacteria/100 ml were detected in one water source of drinkable well water compared with its water tankers. Also there were high number of PA which represent 1170 bacteria/100 ml in one water tanker of non-drinkable well water. Bari *et al.*, (2007) showed lower PA contamination (4%) from wells than that reported in our study. Geldreich (1996) found that PA was widely distributed in nature and most prevalent opportunistic pathogen isolated from the water samples.

P. aeruginosa is part of a large group of free-living bacteria that are ubiquitous in the environment. This organism is often found in natural waters such as lakes and rivers in concentrations of 10/100 mL to >1,000/100 mL. However, it is not often found in drinking water. Usually it is found in 2% of samples, or less, and at concentrations up to 2,300 mL(-1) or more often at 3-4 CFU/mL. Its occurrence in drinking water is probably related more to its ability to colonize biofilms in plumbing fixtures (i.e., faucets, showerheads, etc.) than its presence in the distribution system or treated drinking water (Mena and Gerba, 2009).

Trautmann *et al.*, (2005) in between 1998 and 2005 showed that 9.7% and 68.1% of randomly taken tap water samples on different types of ICUs were positive for PA, and between 14.2% and 50% of infection/colonization episodes in patients were due to genotypes found in ICU water. Although much has been written about biofilms in the drinking water industry, very little has been reported regarding the role of PA in biofilms. Tap water appears to be a significant route of transmission in hospitals, from colonization of plumbing fixtures (Mena and Gerba, 2009). Outbreaks have been reported from exposure to PA in swimming pools and water slides (Mena and Gerba, 2009).

The bacteriological content of water in large dispensers (coolers) and from the 20 liter supply bottles had *P. aeruginosa* in 25% of samples from the large supply bottles and also in 24% of water specimens from the actual coolers. A further 21.6% of 162 specimens from the coolers yielded *P. aeruginosa* (Baumgartner and Grand, 2006).

Enterococcus faecalis

In the present study, *enterococcus faecalis* (*E. faecalis*) was isolated from drinkable and non-drinkable water samples (8.3% and 16.7%) respectively. Ahmed *et al.*, (2005) examined of 12 water samples collected from different wells in Egypt and showed that bacterial contamination *Staphylococcus aureus* (22.22%), *Staphylococcus epidermidis* (11.11%), *Enterococcus faecalis* (11.11%), *Bacillus Cereus* (55.56%), *Yersinia enterocolitica* (37.5%), *Klebsiella pneumoniae* (18.75%), *Pseudomonas aeruginosa* (12.5%), *Escherichia coli* (6.25%), *Enterobacter agglomerans* (6.25%) and *Citrobacter freundii* (6.25%). These findings were similar to the current results in isolation of *P. aeruginosa*, *E. coli* and *E. faecalis*. The differences in results may be to several reasons including geographical differences, types of collection method, number of microorganisms which were isolated and type of water samples.

Our results revealed that *E. faecalis* was found in tankers which was the lowest percentage of bacterial contamination. In contrast to our results, Malakauskas *et al.*, (2007) found *E. faecalis* (23.4%) contamination in may wells. Abdelkarem and Hassan (2000) found that no *E. faecalis* was detected in wells.

In the developing world, 90% of all wastewater still goes untreated into local rivers and streams. About 50 countries, with roughly a third of the world's population, suffer from medium or high water stress, and 17 of these extract more water annually than is recharged through their natural water cycles. *Enterococcus faecalis* not only affects surface freshwater sources (rivers and lakes), but it also degrades groundwater resources (UNEP International Environment, 2002).

Failing home septic systems can allow coliforms in the effluent to flow into the water table,

aquifers, drainage ditches and nearby surface waters. Sewage connections that are connected to storm drain pipes can also allow human sewage into surface waters. Some older industrial cities in USA use a combined sewer system to handle waste. A combined sewer carries both domestic sewage and storm water. During high rainfall periods, a combined sewer can become overloaded and overflow to a nearby stream or river, bypassing treatment (Walkerton, 2011).

Poor quality of water is due to contamination by microorganisms of human or animal origin (Ratajczak *et al.*, 2010). Total coliforms, faecal coliforms, *E. coli* and enterococci are commonly used microbial indicators of water quality (Davis *et al.*, 2005). Several studies of both recreational and drinking water samples suggested that enterococci are more relevant indicators of faecal contamination than faecal coliforms and *E. coli* (Grammenou *et al.*, 2006; Kinzelman *et al.*, 2003).

Approximately 13% of surface waters in USA do not meet designated use criteria because of high densities of faecal indicator bacteria ("Microbial Source Tracking Guide Document" 2005). Despite the uncertainty of the effects of animal faecal contamination of ambient waters to human health, microbiological contamination of recreational waters from human faeces is regarded as a greater risk to human health as they are more likely to contain human-specific pathogens. *E. coli* and Enterococci are considered to have a higher correlation with outbreaks of swimming-associated gastroenteritis than total and faecal coliforms (Wikipedia, 2011).

Twelve water sources (9 wells, 3 taps) and 15 latrines were identified and used by 444 inhabitants. Well and tap water showed heavy faecal contamination with more than 1000 CFU/100 ml. The contamination of drinking water in Bissau due to poor construction, maintenance and improper use ten years after the civil war, demonstrates the need to allocate resources after conflicts in the area of water and sanitation (Colombatti *et al.*, 2009). A total of 300 water samples were collected from 20 different drinking water sources in Kamalapur, Dhaka city from August 2004 to January 2005. The

level of faecal contamination was estimated using measurements of faecal indicator bacteria (total coliforms, faecal coliforms and faecal streptococci). The unacceptable level of contamination of total coliforms (TC), faecal coliforms (FC) and faecal streptococci (FS) ranged from 23.8% to 95.2%, 28.6% to 95.2% and 33.3% to 90.0%, respectively (Sirajul Islam *et al.*, 2007).

Copeland *et al.*, (2009) measured faecal contamination in 231 randomly primary drinking water samples from selected households. Risk for contamination was compared across source and storage types. A third of samples (30.3%) was contaminated; the source with the highest frequency of contamination was well water (23/24: 95.8%). For tap water, the type of storage had a significant effect on the susceptibility to contamination. The observed pattern of contamination demonstrated the relative potential contributions of both source and storage.

Contamination of water samples with *Aspergillus* spp

The recorded results here showed that *Aspergillus* spp. contamination in water tankers and water well source. Warris *et al.*, (2001) reported that 21% of drinking water samples have been contaminated with *Aspergillus* spp. which were higher than that in our obtained results. *Aspergillus* spp. were recorded by Anaissie *et al.*, (2002) to be more as 70% of all drinking water samples examined. In comparing to this study, our findings were lower contamination. *Aspergillus* spp. are opportunistic and can cause health problems in immunocompromised patients (Graybill, 2001).

In Brazil, 50 people died due to algal toxins in water used for haemodialysis in 1996. Toxins can attack the liver, the nervous system or irritate skin, yet very few of these toxins have been isolated and characterized. Taste and odour problems in potable water are increasing worldwide and are produced by microorganisms such as bacteria and fungi (The National Water Research Institute, Canada, 2001).

A total of 197 hot and cold water samples were collected from the main water supply lines and from the taps at three different hospital sites of

the University Hospital of Liège. Filamentous fungi were recovered from 55% and 50% of the main water distribution system and tap water samples, respectively, with a mean of 3.5 and 1.5 colony forming units per 500 ml water. *Aspergillus* spp. were recovered from 6% of the samples of the water distribution system and *A. fumigatus* was the most frequently recovered species (66.6%). *Fusarium* spp. was predominant at one site, where it was found in 28% of tap water samples (Hayette *et al.*, 2010).

Two hundred and forty water samples were collected from four university hospitals. 77.5% were positive for fungal growth. *Aspergillus* (29.7%), *Cladosporium* (26.7%) and *Penicillium* (23.9%) were the most common isolated. Among *Aspergillus* species, *A. flavus* had the highest frequency. Highest colony counts were found in autumn. *Aspergillus* predominated in autumn, *Cladosporium* in winter and spring and *Penicillium* in summer. This results showed that hospital water should be considered as a potential reservoir of fungi particularly *Aspergillus* (Hedayati *et al.*, 2011).

Waterborne fungi have been suspected as a source for allergic reaction in sensitive individuals and they may contribute to produce mycotoxins in water (Hageskal *et al.*, 2009). Therefore, study on fungal contamination of water distribution system especially hospitals water has been an interesting subject for many investigators from different countries in the past decade (Hageskal *et al.*, 2006; Kanzler *et al.*, 2008; Pires-Goncalves *et al.*, 2008; Hayette *et al.*, 2010). Gottlich *et al.*, (2002) reported a mean positivity of 26.6% in a Belgian university hospital and groundwater-derived drinking water from water supplies in Germany.

Contamination of water samples with *Penicillium* spp

Our results indicated that *Penicillium* spp. were found in desalinated water and private desalinated water samples with 100% and 50% respectively. Desalinated water tankers supplies water to private desalinated water stations, therefore, *Penicillium* spp. were detected in both sources. Kanzler *et al.*, (2007) isolated fungi from 38 water wells samples as *Cladosporium* spp.

(74.6%), basidiomycetes (56.4%) and *Penicillium* spp. (48.7%). This study suggested that drinking water can be a reservoir for opportunistic fungal pathogens. These pathogens are naturally found in the environment and are not usually regarded as pathogens. However, they can cause disease in human with impaired defence mechanisms like the elderly or young patients with burns, immunosuppressive therapy patient (Hussain *et al.*, 2001).

Surface water contamination occurs as a result of direct runoff from waste sites to streams, lakes and wetlands, and indirectly as contaminated groundwater discharges to surface waters. The contamination of groundwater is different from surface water contamination. Because we cannot observe groundwater, we typically discover that the groundwater is contaminated once a well or surface water body becomes contaminated. Surface water contamination occurs quickly and can be stopped at the source. However, groundwater contamination may commence years after the waste source is in place. The slow release rate causes it to take years to thousands of years to move through the groundwater flow regime, and groundwater can be difficult, if not impossible to remediate, and prohibitively costly to remediate. Ultimately all contaminated groundwater will discharge to surface water. Thus, should serious groundwater contamination occur, the destruction of drinking water supplies and aquatic ecosystems occurs for decades to hundreds of years (Coote and Gregorich, 2000).

Effects on human civilization

Water fit for human consumption is called drinking water or potable water. Water is made fit for drinking by filtration, distillation, or by a range of other methods. Poor water quality and bad sanitation are deadly; about five million deaths a year are caused by polluted drinking water. WHO (2010) estimates that safe water could prevent 1.4 million child deaths from diarrhoea each year.

The contamination of drinking water sources with microbial pathogens is an on-going problem. More than three million people die every year from water-related disease and 43% of water-related deaths are due to diarrhoea (WHO, 2008).

The majority of diseases are infectious in nature caused by bacteria, fungi, viruses and parasites, excreted in human faeces which may lead to contaminate water supplies (Tambekar and Hirulkar, 2007).

Well water is one of the water sources which can be used for population purposes. In the current study, one (16.7%) out of all wells of drinkable well water is clean. Therefore, the remaining wells found to be contaminated with bacterial and fungal pathogens. In Ontario, there are an estimated 500,000 wells which about 10% - 34% of these wells were clean and there were no contamination (Goss *et al.*, 1998). An investigation carried out in Nigeria found that all water samples collected from 15 wells were bacterial contaminated (Olabisi *et al.*, 2008). Same study estimated 20 colony counts per 100 ml as a maximum value, while the present study found that the maximum value of colony counts was 117 per 100 ml.

From the obtained results in our study we can conclude that: (1) Certain wells and certain water tankers were found to be contaminated with different microbial pathogens, bacteria and fungi. (2) *Penicillium* spp. was detected only in desalinated water and private desalinated water before filtration process. (3) The suitable water for drinking is the private desalinated water because they come under different treatment processes like filtration.

Recommendations

Safe water and sanitation pose universal challenges for public health as:

- 1 Periodical monitoring of water sources for pollutants (chemical & microbial).
- 2 Periodical testing of water tankers for their microbial contamination. The risk of microbial contamination in tanks can be reduced by

several well-known practices. These include the installation of first flush devices, cleaning gutters, both of which are designed to reduce the build-up of potential contaminants and the use of filtration to remove potential contaminants before use

- 3 Enhanced funding is needed to validate newer molecular detection tools, understand the ecology of pathogens in aquatic ecosystems, better predict disease outbreaks, and improve emergency responses. A preventive approach to pathogen pollution should be taken by developing countries in the form of a source water protection program for all major freshwater sources.
- 4 The identification and control of threats posed by waterborne pathogens will also require effective pathogen detection techniques. The need to develop, evaluate and validate newer molecular tools for pathogen detection such as PCR techniques and DNA microarrays. Rapid advances in fields such as genomics offer the potential to develop improved pathogen detection tools.
- 5 Encourage infrastructure planning, including technological advances, to ensure that improved treatment and environmental protection measures are not diminished by development or population growth.
- 6 Programs are needed to assess the effects of aerial emissions on drinking water quality.
- 7 An improved understanding is needed of methods for assessment and risk analysis of the cumulative effects of agricultural, forestry and other land use activities (e.g., ore, oil and gas exploration) as well as point-source inputs (e.g., municipal and industrial discharges) on surface and ground waters.

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